Total Synthesis of Sialylgalactosylgloboside: Stage-Specific **Embryonic Antigen 4**

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A versatile total synthesis of sialylgalactosylgloboside (SGG, 1), carrying the stage-specific embryonic antigen 4 (SSEA-4) is reported, illustrating a more general strategy for the synthesis of complex globo-series glycosphingolipids. Starting from readily available building blocks 7, 8, and 10, two different approaches to the synthesis of the key tetrasaccharide **6** have been developed in a highly convergent manner. Further glycosylations with galactosyl trichloroacetimidate (5) and sialyl phosphite (2) donors successively afforded the penta- and hexasaccharides 3 and 11. The latter was finally converted into the target molecule (SGG, 1) with the help of a azidosphingosine glycosylation procedure, favored in this case by the stereocontrolling properties of the 2a-O-pivaloyl protecting group. Valuable intermediates **6** and **3**, having the oligosaccharidic skeletons of Gb_4 and Gb₅ (SSEA-3), respectively, were obtained in the course of the synthesis.

Glycosphingolipids (GLS's) of the globo series are membrane-associated antigens possessing as structural feature a Gal α 1 \rightarrow 4Gal (galabiose-like) linkage which is responsible for their specific roles in recognition and cell differentiation. They are recognized by proteins of pathogenic Escherichia coli in human epithelial cells of the urinary tract.¹ They are also receptors for Shiga toxin² and verotoxin (Shiga-like toxin)³ and constitute the basis of the P-blood group system.⁴ Many of them have been described as tumor-associated antigens.⁵ Sialylgalactosylgloboside (SGG, 1), a ganglioside belonging to this series, was first isolated in 1983 from chicken pectoral muscles.⁶ In the same year, Solter et al.⁷ identified SGG with a developmentally regulated antigen, the stagespecific embryonic antigen 4 (SSEA-4), and found SSEA-3 (previously found to be expressed as Gb₅)⁸ and SSEA-4 to be epitopes of this unique globo-series ganglioside isolated from human teratocarcinoma cells. More recent studies have shown that *globo*-series antigens SEEA-3 and SSEA-4 are expressed in different forms on human and murine teratocarcinoma cells9 and are a hallmark

⁹ Abstract published in Advance ACS Abstracts, August 15, 1996. Lanne, B.; Olsson, B.-M.; Jovalp, P.-Å.; Angström, J.; Linder, H.; Marklund, B.-I.; Bergström, J.; Karlsson K.-A. J. Biol. Chem. 1995, 270, 9017, and references cited therein. Our synthetic SGG is currently being used in this laboratory for further investigations on this topic.

(2) Lindberg, A. A.; Brown, J. E.; Strömbreg, N.; Westling-Ryd, M.; Schultz, J. E.; Karlsson, K. A. *J. Biol. Chem.* **1987**, *262*, 1779.

(3) Linwood, C. A.; Law, H.; Richardson, S.; Petric, M.; Brunton, J. L.; De Grandis, S.; Karmali, M. *J. Biol. Chem.* **1987**, *262*, 8834.

(4) (a) Naiki, M.; Marcus, D. M. Biochem. Biophys. Res. Commun.

(b) Raistak, A., Millindi, J. G., Obster Wirk, E., Ota, E. J., Ota, E. J., Schemberg, D. A. *Int. J. Cancer* **1991**, *49*, 837.
(6) Chien, J.-L.; Hogan, E. L. *J. Biol. Chem.* **1983**, *258*, 10727.
(7) Kannagi, R.; Cochran, N. A.; Ishigami, F.; Hakomori, S.-i.; Andrews, P. W.; Knowles, B. B.; Solter, D. *EMBO J.* **1983**, *2*, 2355.

(8) (a) Kannagi, R.; Levery, S. B.; Ishigami, F.; Hakomori, S.-i., Shevinski, L. H.; Knowles, B. B.; Solter, D. *J. Biol. Chem.* **1983**, *258*, 8934. (b) Shevinski, L. H.; Knowles, B. B., Damjanov, I.; Solter, D. *Cell* **1982**, *30*, 697.

(9) Krupnick, J. G.; Damjanov, I.; Damjanov, A.; Zhu, Z. M.: Fenderson, B. A. *Int. J. Cancer* **1994**, *59*, 692.

of the former.¹⁰ SSEA-4 has also been reported to bind human parvovirus B19 capsids.¹¹

The great interest in this family of GLS's and the difficulties for their isolation in sufficient quantities from natural sources has stimulated interest in their chemical synthesis. Table 1 summarizes the most significant contributions that have appeared up to now in this field.

We describe herein a general strategy for the synthesis of complex globo-series glycosphingolipids, as illustrated for the detailed synthesis of SGG, carrying the SSEA-4 antigen, and including the Gb4 and Gb5 oligosaccharidic skeletons as valuable intermediates. A preliminary account of this work¹⁷ and a different approach to the synthesis of SGG have recently appeared.¹⁸

Results and Discussion

The retrosynthetic view of the target compound 1 outlined in Scheme 1 suggests six convenient disconnections designed in such a way that useful intermediates would be obtained within the course of the synthesis. Thus, the access to the target molecule is visualized from compound 3, having the oligosaccharidic skeleton of galactosylgloboside (Gb5, SSEA-3) and known building blocks 2^{19} and 4^{20} (disconnections 1–3). Likewise, com-

(12) (a) Shapiro, D.; Acher, A. J.; *Chem. Phys. Lipids* **1978**, *22*, 197.
(b) Koike, K.; Sugimoto, M.; Sato, S.; Ito, Y.; Nakahara, Y.; Ogawa, T. Carbohydr. Res. **1987**, *163*, 189. (c) Nicolaou, K. C.; Caulfield, T.; Kataoka, H.; Kumazawa, T. J. Am. Chem. Soc. **1988**, *110*, 7910. (d) (13) Leontein, K.; Nilsson, M.; Norberg, T. Carbohydr. Res. 1985,

144, 231.

(14) Magnusson, G.; Nilsson, U.; Ray, A. K.; Taylor, K. G. ACS Symp. Ser. **1993**, *519*, 92, and references therein.

(15) Nunomura, S.; Ogawa, T. Tetrahedron Lett. 1988, 29, 5681.

 (16) (a) Bilodeau, M. T.; Park, T. K.; Hu, S.; Randolph, J. T.;
 Danishefsky, S. J.; Livingston, P. O.; Zhang, S. J. Am. Chem. Soc. 1995, 117, 7840. (b) Lassaletta, J. M.; Schmidt, R. R. Liebigs Ann. Chem. submitted.

(17) Lassaletta, J. M.; Schmidt, R. R. Tetrahedron Lett. 1995, 36, 4209.

(18) Ishida, H.; Miyawaki, R.; Kiso, M.; Hasegawa, A. J. Carbohydr.

Chem 1996, 15, 163. (19) Martin, T. J.; Brescello, R.; Toepfer, A.; Schmidt, R. R. Glyco-conjugate J. 1993, 16, and references cited therein.

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⁽¹⁰⁾ Wenk, J.; Andrews, P. W., Casper, J.; Hata, J. I.; Pera, M. F.; Vonkeitz, A., Damjanov, I.; Fenderson, B. A. Int. J. Cancer 1994, 58, 108.

⁽¹¹⁾ Cooling, L. L. V.; Koerner, T. A. W.; Naides, S. J. J. Infect. Dis. 1995, 172, 1198.





pound **3** should be accessible starting from tetrasaccharide **6** (Gb4) and the readily available galactosyl donor 5^{21} (disconnection 4). A convergent retrosynthetic view for the former leads to compound **10** as lactose building block and disaccharide donor **9** (disconnection 5); evident disconnection (6) of the latter is proposed from galactose acceptor **8** and azidogalactose donor **7**.

Two different approaches for the synthesis of the globotetraose skeleton **6** have been tested (Scheme 2). The choice of an appropriate AB (lactose) building block was made considering the following requirements: (i) the low nucleophilicity of 4b-OH should be enhanced by benzyl-like protecting groups around this position; (b) a sterically demanding β -stereocontrolling group should be placed at the 2a-position, granting the formation of β -anomers only and avoiding the formation of orthoesters

at the azidosphingosine glycosylation step.²² Compound **10**,²³ fulfilling these requisites, was chosen for both approaches. For the first synthesis of the CD building block **9**, the known 3-*O*-monochloroacetyl-protected azidogalactosyl donor **7a**²⁴ was reacted with acceptor **8d**. The latter was synthesized starting from allyl galactoside **8a**²⁵ in three steps: selective *O*-benzoylation (BzCN, Et₃N), benzylation under neutral conditions (BnOTf),²⁶ and saponification of the 3-*O*-benzoyl group. The stereochemistry of the glycosylation reaction was β -directed by

^{(20) (}a) Schmidt, R. R.; Zimmermann, P.; *Tetrahedron Lett.* **1986**, *27*, 481. (b) Zimmermann, P.; Bommer, R.; Bär, T.; Schmidt, R. R. J. Carbohydr. Chem. **1988**, *7*, 435.

⁽²¹⁾ Greilich. U.; Zimmermann, P.; Jung, K.-H.; Schmidt, R. R. Liebigs Ann. Chem. 1993, 859.

^{(22) (}a) Schmidt, R. R.; Zimmermann, P. Angew. Chem. Int. Ed. Engl. **1986**, *25*, 725, and references therein. (b) Kunz, H.; Harreus, A. Liebigs Ann. Chem. **1982**, 41.

⁽²³⁾ This compound was first synthesized by Ogawa's group: Sato, S.; Nunomura, S.; Nakano, T.; Ito, Y.; Ogawa, T. *Tetrahedron Lett.* **1988**, *29*, 4097. The low overall yield and length (14 steps) of the synthesis, however, prompted us to develop a shorter, more efficient synthesis of this compound: Lassaletta, J. M.; Schmidt, R. R. *Synlett* **1995**, 925.

^{(24) (}a) Grundler, G.; Schmidt, R. R. *Liebigs Ann. Chem.* 1984, 1826.
(b) Rademann, J. Diplomarbeit, University of Konstanz, 1994.

⁽²⁵⁾ This compound was obtained in good yield by direct anomeric *O*-alkylation of 4,6-*O*-benzylidenegalactopyranose (see Experimental Section).



^{*a*} Reagents: (a) BzCN, Et₃N (75%); (b) BnOTf (81%); (c) K₂CO₃, MeOH/H₂O (85%); (d) TMSOTf, CH₃CN; (e) PdCl₂, NaOAc, AcOH/ H₂O (71%); (f) NaOMe, MeOH (quant); (g) Cl₃CCN, DBU (76%); (h) TMSOTf, Et₂O, IP (62%); (i) HCl, MeOH/H₂O (51%); (j) (*j*) TBAF, AcOH (92%); (*ii*) CCl₃CN, DBU (88%); (k) TMSOTf, CH₃CN (84%, *β*); (l) BnBr, Ag₂O, DMF, 4 Å (81%); (m) PdCl₂, NaOAc, AcOH/H₂O (86%); (n) CCl₃CN, DBU (93%); (o) TMSOTf, IP (63%, $\alpha:\beta = 4:1$); (p) NH₃/MeOH (quant).

using conditions (-40 °C, CH₃CN) in which the "nitrile effect" operates;²⁷ **9a** was isolated in 88% yield and no α -product could be detected. The allyloxy group of **9a** could not be isomerized under standard conditions (i.e., Wilkinson's catalyst in a protic solvent in the presence of a DBU-like base) because partial cleavage of the labile monochloroacetyl (MCA) group occurred (\rightarrow **9b**). Use of the buffered Ogawa's reagent²⁸ (PdCl₂/AcOH/NaOAc),

however, afforded directly the 1-*O*-deprotected disaccharide **9c** in good yield, conditions being mild enough to keep the acid-labile benzylidene protecting groups. The transformation of **9c** into the corresponding trichloroacetimidate donor again presented problems due to the lability of the MCA group, which did not survive the basic conditions required. To overcome this problem, **9c** was first quantitatively deprotected to diol **9d** and subsequently transformed into the bis-trichloroacetimidate **9e**. Reaction of the latter with acceptor **10** furnished, with the help of the inverse procedure (IP),²⁹ the desired tetrasaccharide **6a** in a yield of 62%, acceptable for this system. HCl-mediated hydrolysis of the 3d-*O*-trichloroacetimidoyl group afforded acceptor **6b** in moderate (51%) yield.

The general importance of the ABCD tetrasaccharide, having the core structure of globoside Gb4 and being the key for all higher globosides, merits an improved synthesis. Therefore, a second approach was designed to overcome the limitations referred to above. In this case, the known tert-butyldimethylsilyl 2-azido-4,6-O-benzylidene- β -D-galactopyranoside **7b**^{24a} was converted into donor 7c by treatment with (i) TBAF and (ii) Cl₃CCN/ DBU, without isolation of the intermediate hemiacetal. Reaction of 7c with diol 8a proceeded regio- and stereoselectively, again taking advantage of the "nitrile effect"27 and the much higher reactivity of 3-OH than 2-OH in the latter. Thus, the desired disaccharide 9f was obtained as a single isomer and then transformed into trichloroacetimidate CD donor 9i by benzylation (Ag₂O/ BnBr, \rightarrow **9g**), allyl cleavage (PdCl₂/AcOH/NaOAc, \rightarrow **9h**) and imidate formation under established conditions. Reaction of this compound with acceptor 10, again applying the inverse procedure,²⁹ afforded tetrasaccharide **6c** (63%, α : β = 4:1). Selective removal of the 3d-*O*acetyl group to obtain 6b was then accomplished quantitatively by means of methanolic ammonia, keeping the 2a-O-pivaloyl group untouched. This material proved to be identical with product 6b obtained from 6a. The second approach, however, proved to be more efficient, since the synthesis required fewer steps, and the overall yield was higher.

For the transformation of this ABCD fragment 6b into the target **1** (Scheme 3), β -galactosylation was first achieved by using trichloroacetimidate donor 5 under established conditions, yielding the globopentaose derivative **3a** in almost quantitative yield. This material was converted into the ABCDE acceptor **3b** again using NH₃/ MeOH for selective de-O-acylation. It is known¹⁹ that sterically demanding sialyl donors can be attached in good yields to the 3 position of 2,3,4- or 2,3-O-unprotected galactose residues. Therefore, compound **3b** reacted with phosphite donor 2,14 affording an easily separable mixture **11a** α /**11a** β in good yield (62%, $\alpha:\beta = 4:1$).³⁰ The stereochemistry of both α and β sialylglycosides was assigned on the basis of empirical rules.³¹ Compound **11a** was then converted into trichloroacetimidate 11d in three convenient and high-yielding steps: (i) simultaneous hydrogenolytic cleavage of benzyl and benzylidene protecting groups and reduction of the 2d-azido group, best

⁽²⁶⁾ Lemieux, R. U.; Kondo, T. Carbohydr. Res. **1974**, 35, C4. Other standard conditions (NaH/BnBr or $Ag_2O/BnBr$) led to benzoyl migration.

⁽²⁷⁾ Schmidt, R. R.; Behrendt, M.; Toepfer, A. *Synlett* **1990**, 694. (28) Ogawa, T.; Nakabayashi, S. *Carbohydr. Res.* **1981**, *93*, C1.

⁽²⁹⁾ Schmidt, R. R.; Toepfer, A. *Tetrahedron Lett.* **1991**, *32*, 3353. (30) Though the phosphite methodology (see ref 19) usually yields virtually complete α -selectivities, other rather unpredictable results have been observed for related systems. This result, however, can be considered as acceptable due to the high yield obtained.

considered as acceptable due to the high yield obtained. (31) a) δ 3-Heq(α) > δ 3-Heq(β): Dabrowski, U.; Friebolin, H.; Brossmer, R.; Supp, M. *Tetrahedron Lett.* **1979**, 4637. (b) δ 4-H(α) < δ 4-H(β): Paulsen, H.; Tietz, H. *Carbohydr. Res.* **1984**, *125*, 47.

Scheme 3^a



^{*a*} Reagents: (a) TMSOTf (94%); (b) NH₃/MeOH (quant); (c) TMSOTf, CH₃CN (62%, α: β = 3.5:1); (d) (*i*) Pd(OH)₂/C, H₂, MeOH, (*ii*) Ac₂O/Py (85%, α: β = 1:1); (e) N₂H₄·AcOH, DMF (94%); (f) CCl₃CN, DBU (93%); (g) TMSOTf (72%); (h) (*i*) H₂S, Py/H₂O, (*ii*) C₁₅H₃₁COOH, WSC (87%); (j) (*i*) NaOMe/MeOH, (*ii*) KOH/H₂O, (*iii*) IRA 120 (H⁺) (quant).

achieved by using freshly prepared Pearlman catalyst,32 followed by peracetylation of the crude product (\rightarrow **11b**, 85% of a 1:1 mixture of α - and β -acetates); (ii) regioselective removal of the anomeric 1-O-acetyl group by means of hydrazinium acetate³³ in DMF (\rightarrow **11c**, 94%); and iii) imidate formation under established conditions $(\rightarrow 11d, 93\%)$. The final stages of the synthesis, i.e., the introduction of the ceramide aglycon, were accomplished by application of the azidosphingosine glycosylation procedure.²⁰ The importance of the pivaloyl protecting group chosen for the 2a position of 11d was here demonstrated by the excellent yield (72%) observed for the TMSOTf-promoted β -glycosylation of azidosphingosine derivative 4, which yielded the corresponding glycoside 12a; no orthoester byproduct was detected in the reaction mixture. This azido compound was then reduced with H₂S in pyridine/water without acyl migration, and the crude product was immediately treated with palmitic acid in the presence of N-[3-(N,N-dimethylamino)propyl]-N-ethylcarbodiimide hydrochloride (water soluble carbodiimide, WSC) as condensing agent to provide the protected glycosyl ceramide 12b in 87% yield. Finally, the target 1 (SGG) was obtained quantitatively from the latter by methanolysis (NaOMe/MeOH) of all O-acyl protecting groups and subsequent hydrolysis (aqueous KOH) of the methyl ester moiety. The ¹H NMR data of this compound are in full agreement with those reported for the natural sample.^{8a,34}

Conclusions

In summary, a versatile strategy has been described for the synthesis of complex globo-series glycolipids, based on powerful synthetic tools such as the trichloroacetimidate glycosylation procedure, the recently developed phosphite method for sialylation,¹⁹ the azidosphingosine-based protocol for glycosphingolipid synthesis, and the stereocontrolling properties of a neighbor pivaloyl protecting group. An advantage of this strategy is the construction of the difficult galabiose-type $\alpha(1 \rightarrow 4)$ glycosidic linkage in the first stages of the synthesis, thus avoiding this connection later during the synthesis. It should be also stressed that both **3b** and **6b**, bearing appropriate protecting groups, are not only precursors for the synthesis of Gb₄ and Gb₅ globosides but also for further elongation to several higher globosides other than the target SGG. Compound 6b, for instance, could be alternatively glycosylated with an α -galactosamine donor for the synthesis of the Forssman antigen. Likewise, the considerable difference in reactivity between the 3d-OH

⁽³²⁾ Fieser, L. F.; Fieser, M. In *Reagents for Organic Synthesis*; John Wiley and Sons Inc.: New York, 1967; Vol. 1, p 782. The age of the catalyst was found to be of importance; in our hands, the use of old catalysts led to complex mixtures containing the desired product in low yield.

⁽³³⁾ Excoffier, G.; Gagnaire, G.; Utille, J.-P. *Carbohydr. Res.* **1975**, *39*, 368.

⁽³⁴⁾ The molecular formula and the $^1\mathrm{H}$ NMR data reported for SGG in ref 18 are erroneous.

and 2d-OH of 3b can be used for the synthesis of Globo-H¹⁶ and Globo-A structures. Additionally, the lysoglycosphingolipid obtained after reduction of 12a is another interesting intermediate, since it may have different biological properties to that of the corresponding ceramide derivative.35

Experimental Section

Melting points were measured in a metal block and are uncorrected. Thin layer chromatography (TLC) was performed on silica gel precoated plates. Purifications of the products were carried out by flash chromatography. The light petro-leum ether used had boiling range 35-65 °C. ¹H NMR spectra were recorded at 250 MHz, using chloroform-d as solvent and tetramethylsilane as internal standard, unless noted otherwise.

Starting Materials. 4,6-O-Benzylidene- α , β -D-galactopyranose, 36 5-acetamido-4,7,8,9-tetra-O-acetyl-3,5-dideoxy-1-(methoxycarbonyl)-D-glycero-a-D-galacto-2-nonulopyranosyl diethyl phosphite¹⁹ (2), (2S,3R,4E)-2-azido-3-(benzoyloxy)-4-octa-decen-1-ol²⁰ (4), 2,3-di-O-acetyl-4,6-O-benzylidene- α -D-galactopyranosyl trichloroacetimidate²¹ (5), 2-azido-4,6-O-benzylidene-3-O-chloroacetyl-2-deoxy-α-D-galactopyranosyl trichloroacetimidate²⁴ (7a), tert-butyldimethylsilyl 2-azido-4,6-O-benzylidene- β -D-galactopyranoside^{24a} (7b), and benzyl 3,6-di-O-benzyl-4-O-(2,3,6-tri-O-benzyl- β -D-galactopyranosyl)-2-O-pivaloyl- β -Dglucopyranoside²³ (10) were prepared according to literature procedures.

Allyl 4,6-*O*-Benzylidene- β -D-galactopyranoside (8a). To a stirred solution of 4,6-*O*-benzylidene- α , β -D-galactopyranose (33.8 g, 126 mmol) and AllBr (17 mL, 200 mmol) in 1:1 dry THF/DMF (300 mL) was added NaH (3.56 g, 141 mmol) in small portions. After 24 h MeOH (10 mL) was added and the mixture stirred for another 30 min. The solvent was then removed in vacuo and the residue extracted with acetone and purified by flash chromatography (3:1 toluene-acetone). Allyl 4,6-*O*-benzylidene- α -D-galactopyranoside (7.3 g, 13%) eluted first: mp 118–120 °C (lit.³⁷ mp 115–117 °C); [α]^{rt}_D +73.6° (*c* 1, CHCl₃) [lit.³⁷ [α]^{rt}_D +121 ° (*c* 1.9, CHCl₃)]. **8a** eluted second (22.8 g, 59%): mp 174 °C; $[\alpha]^{rt}_{D}$ –36.4° (*c* 1, CHCl₃); TLC (12:1 CH₂Cl₂-MeOH) R_f 0.57; ¹H-NMR δ 7.3-7.5 (m, 5H), 6.02-5.86 (m, 1H), 5.52 (s, 1H), 5.35-5.17 (m, 2H), 4.46-4.38 (m, 1H), 4.35-3.60 (m, 7H), 3.44 (br s, 1H), 2.65 (br s, 2H). Anal. Calcd for C₁₆H₂₀O₆·¹/₄H₂O: C, 61.43; H, 6.60. Found: C, 61.44; H. 6.49.

Allyl 3-O-Benzoyl-4,6-O-benzylidene-β-D-galactopyra**noside (8b).** To a stirred suspension of **8a** (18 g, 58 mmol) and BzCN (8.0 g, 1.03 eq) in dry CH₃CN (150 mL) was added Et₃N (10 drops). After 30 min MeOH (10 mL) was added and the mixture stirred for another 15 min. The solvent was then removed in vacuo and the residue dissolved in MeOH and concentrated two times. Crystallization from MeOH and flash chromatography [2:1 toluene-ethyl acetate (EA)] of the mother liquors yielded unreacted material (3 g) and **8b** (17.7 g, 75%): mp 177 °C; $[\alpha]^{rt}_{D}$ +76.8° (c 1, CHCl₃); TLC (4:1 tolueneacetone) Rf 0.38; ¹H-NMR & 7.2-8.1 (m, 10H), 6.03-5.87 (m, 1H), 5.50 (s, 1H), 5.19–5.36 (m, 2H), 5.14 (dd, J = 3.7, 10.3 Hz, 1H), 4.51-4.05 (m, 7H), 3.57 (br s, 1H), 2.0 (br s, 1H). Anal. Calcd for C₂₃H₂₄O₇: C, 66.96; H, 5.87. Found: C, 66.86; H. 5.87.

Allyl 3-O-Benzoyl-2-O-benzyl-4,6-O-benzylidene-β-Dgalactopyranoside (8c). To a solution of Tf_2O (14.26 mL, 85.6 mmol) in dry CH_2Cl_2 (40 mL) was added dropwise at -75°C a solution of benzyl alcohol (9.0 mL, 85.6 mmol) and 2,6di-tert-butylpyridine (18.8 mL, 85.6 mmol) in dry CH₂Cl₂ (70 mL) for over 30 min under an argon atmosphere. After stirring for 20 min, a solution of compound 8b (11.6 g, 28.2 mmol) and 2,6-di-tert-butylpyridine (12.54 mL, 57 mmol) in CH₂Cl₂ (70 mL) was added dropwise (20 min) and the temperature was allowed to rise up to -40 °C. After 16 h, pyridine (20 mL) was added, and the mixture was warmed up to rt, washed with H_2O (3 × 100 mL), dried (MgSO₄), and concentrated *in vacuo*. The 2,6-di-tert-butylpyridine was finally removed by distillation (34 °C/0.4 mbar) and the residue purified by flash chromatography (18:1 toluene-EA) to give unreacted material (1.9 g) and crystalline **8c** (8.64 g, 73%): mp 51 °C; $[\alpha]^{rt}$ +143.8° (c 1, CHCl₃); TLC (3:1 toluene-acetone) R_f 0.47; ¹H-NMR δ 7.1–8.1 (m, 15H), 6.06–5.90 (m, 1H), 5.51 (s, 1H), 5.41-5.22 (m, 2H), 5.20 (dd, J = 3.6, 10.2 Hz, 1H), 4.92 (d, J= 10.1 Hz, 1H), 4.72 (d, J = 10.1 Hz, 1H), 4.61 (d, J = 7.8 Hz, 1H), 4-55-4-03 (m, 6H), 3.56 (br s, 1H). Anal. Calcd for C₃₀H₃₀O₇·¹/₄H₂O: C, 71.06; H, 5.96. Found: C, 70.90; H, 5.99.

Allyl 2-O-Benzyl-4,6-O-benzylidene- β -D-galactopyranoside (8d). To a solution of 8c (11.22 g, 22.3 mmol) in 8:1 MeOH-H₂O was added K₂CO₃ (1M, 2 mL) and the mixture stirred for 7 h at rt. The solvent was then evaporated in vacuo and the residue extracted with acetone and purified by flash chromatography (7:1 toluene-acetone) to give 8d (7.53 g, 85%): mp 69 °C; [α]^{rt}_D +9.3° (c 1, CHCl₃); TLC (7:1 tolueneacetone) $R_f 0.19$; ¹H-NMR δ 7.6–7.2 (m, 10H), 6.05–5.89 (m, 1H), 5.56 (s, 1H), 5.39–5.18 (m, 2H), 5.00 (d, J = 10.1 Hz, 1H), 4.73 (d, J = 10.1 Hz, 1H), 4.6–3.6 (m, 8H), 3.43–3.42 (m, 1H), 2.51 (d, J = 8.0 Hz, 1H). Anal. Calcd for $C_{23}H_{26}O_6$. H₂O: C, 66.33; H, 6.77. Found: C, 66.36; H, 6.51.

Allyl 3-O-(2-Azido-4,6-O-benzylidene-3-O-(chloroacetyl)-2-deoxy-β-D-galactopyranosyl)-2-O-benzyl-4,6-O-benzylidene- β -D-galactopyranoside (9a). To a cooled (-40 °C) solution of $\mathbf{8d}$ (5.33 g, 13.38 mmol) and $\mathbf{7a}$ (10.46 g, 20.07 mmol) in dry CH₃CN (100 mL) was added TMSOTf (2.6 mL, 0.1M in CH₃CN) under an argon atmosphere. After 1 h, NaHCO₃ (s, 0.1 g) and Et₂O (400 mL) were added, and the mixture was washed with saturated NaHCO₃ solution (100 mL) and H₂O (100 mL). The organic layer was dried (MgSO₄) and concentrated in vacuo, and the residue was purified by flash chromatography (7:1 toluene-EA) to afford **9a** (8.85 g, 88%): mp 112 °C; $[\alpha]^{rt}$ +25.4° (*c* 1, CHCl₃); TLC (7:1 toluene– acetone) $R_f 0.50$; ¹H-NMR δ 7.6–7.2 (m, 15H), 6.04–5.88 (m, 1H), 5.59 (s, 1H), 5.48 (s, 1H), 5.37–5.17 (m, 2H), 5.02 (d, J= 10.6 Hz, 1H), 4.97 (d, J = 7.2 Hz, 1H), 4.70 (d, J = 10.6 Hz, 1H), 4.65 (dd, J = 3.6, 10.9 Hz, 1H), 4.53–3.90 (m, 14H), 3.39 (br s, 1H), 3.31 (br s, 1H). Anal. Calcd for C₃₈H₄₀ClN₃O₁₁: C, 60.84; H, 5.37; N, 5.60. Found: C, 60.84; H, 5.55; N, 5.39.

Allyl 3-O-(2-Azido-4,6-O-benzylidene-2-deoxy-β-D-galactopyranosyl)-2-O-benzyl-4,6-O-benzylidene-β-D-galactopyranoside (9b). (a) To a solution of compound 9a (50 mg, 67μ mol) in MeOH (1 mL) was added Et₃N (1 drop), the mixture was stirred for 1 h at rt and concentrated, and the residue purified by flash chromatography (2:1 EA-toluene) to give **9b** (43 mg, 96%): mp 120 °C; [α]^{rt}_D +3.9° (c 1, CHCl₃); TLC (2:1 EA-toluene) R_f 0.29; ¹H-NMR δ 7.6–7.2 (m, 15H), 6.04-5.90 (m, 1H), 5.59 (s, 1H), 5.52 (s, 1H), 5.37-5.17 (m, 2H), 5.00 (d, J = 10.1 Hz, 1H), 4.83 (d, J = 8.0 Hz, 1H), 4.74 (d, J = 10.1 Hz, 1H), 4.55 - 3.27 (m, 15H), 2.60 (d, J = 9.6 Hz, 1H). Anal. Calcd for C₃₆H₃₉N₃O₁₀·H₂O: C, 62.51; H, 5.97; N, 6.07. Found: C, 62.58; H, 5.93; N, 6.00.

(b) A mixture of **9a** (50 mg, 67 µmol), Rh(PPh₃)₃Cl (6 mg, 6.7 μ mol), and 9:1 EtOH–DBU (10 μ L) in 9:1 EtOH–H₂O (1 mL) was stirred at rt for 2 h. The solvent was removed in vacuo and the residue purified by flash chromatography (2:1 EA-toluene) to give 9b (30 mg, 64%), identical with the product described above.

3-O-(2-Azido-4,6-O-benzylidene-3-O-(chloroacetyl)-2deoxy-\beta-D-galactopyranosyl)-2-O-benzyl-4,6-O-benzylidene- α/β -D-galactopyranose (9c). Compound 9a (6.24 g, 8.3 mmol), PdCl₂ (3.4 g, 19.2 mmol), and NaOAc (11.7 g, 125 mmol) were suspended in 9:1 AcOH-H₂O (200 mL) and stirred for 12 h at rt. The mixture was then filtered through Celite, slowly poured on saturated NaHCO₃ solution ($\tilde{1}$ L), and extracted with CH_2Cl_2 (3 × 300 mL). The organic layer was then dried (MgSO₄) and concentrated *in vacuo*, and the residue was purified by flash chromatography (2:1 toluene-EA) to yield **9c** (4.21 g, 71%); [α]^{rt}_D +43.4° (*c* 1, CHCl₃); TLC (2:1 EAtoluene) $R_f 0.55$; ¹H-NMR δ 7.6–7.2 (m, 15H), 5.59 (s, 1H), 5.50 and 5.48 (2 s, 1H), 5.36 (d, J = 2.0 Hz, 0.5H), 5.07 (d, J= 8.0 Hz, 0.5H), 5.0–3.3 (m, 18H). Anal. Calcd for $C_{35}H_{36}$ -

^{(35) (}a) Hannun, Y. A.; Bell, R. M. *Science* **1989**, *243*, 500. (b) Hakomori, S.-i. *J. Biol. Chem.* **1990**, *265*, 18713. (36) Gros, E. G.; Deulofeu, V. *J. Org. Chem* **1964**, *29*, 3647. (37) Gigg, R.; Warren, C. D. *J. Chem. Soc. (C)* **1968**, 1903.

 $ClN_{3}O_{11}\textbf{\cdot}^{1}\!/_{4}H_{2}O:$ C, 58.45; H, 5.19; N, 5.84. Found: C, 58.68; H, 5.24; N, 5.37.

3-*O*-(**2**-Azido-4,6-*O*-benzylidene-2-deoxy-β-D-galactopyranosyl)-2-*O*-benzyl-4,6-*O*-benzylidene-α/β-D-galactopyranose (9d). To a solution of 9c (0.8 g, 1.1 mmol) in MeOH (10 mL) was added NaOMe (0.2 M in MeOH, few drops). After 5 min, ion exchange resin Amberlite IR 120 (H⁺) (0.1 g) was added and the mixture was stirred for another 10 min, filtered, and concentrated *in vacuo* to give pure 9d (0.68 g, quant); $[\alpha]^{rt}_D$ +47.2° (*c* 1, CHCl₃); TLC (5:2 toluene-acetone) R_f 0.26; ¹H-NMR δ 7.6-7.2 (m, 15H), 5.59 and 5.52 (2 s, 1H), 5.57 and 5.53 (2 s, 1H), 5.37 (d, J = 1.9 Hz, 0.7H), 4.96 (d, J = 7.9 Hz, 0.3H), 4.86-3.30 (m, 17H). Anal. Calcd for C₃₃H₃₅N₃O₁₀·¹/₄-H₂O: C, 62.11; H, 5.61; N, 6.58. Found: C, 62.16; H, 5.64; N, 6.39.

3-*O*-(2-Azido-4,6-*O*-benzylidene-2-deoxy-3-*O*-(trichloroacetimidoyl)-β-D-galactopyranosyl)-2-*O*-benzyl-4,6-*O*-benzylidene-α-D-galactopyranosyl Trichloroacetimidate (9e). To a solution of 9d (120 mg, 0.19 mmol) and Cl₃CCN (0.4 mL) in dry CH₂Cl₂ was added DBU (1 drop) and the mixture was stirred for 2 h under an argon atmosphere. The solvent was then removed *in vacuo* and the residue purified by flash chromatography (8:1 toluene-acetone) to yield 9e (130 mg, 76%) as a foam: TLC (8:1 toluene-acetone) R_f 0.31; ¹H-NMR δ 8.57 (s, 1H), 8.46 (s, 1H), 7.2–7.6 (m, 15H), 6.58 (d, J = 2.4 Hz, 1H), 5.60 (s, 1H), 5.55 (s, 1H), 4.93 (d, J = 8.0 Hz, 1H), 4.8–4.0 (m, 6H), 3.91 (br s, 1H), 3.46 (br s, 1H).

Benzyl O-(2-Azido-4,6-O-benzylidene-2-deoxy-3-O-(trichloroacetimidoyl)- β -D-galactopyranosyl)-(1 \rightarrow 3)-O-(2-Obenzyl-4,6-O-benzylidene-α-D-galactopyranosyl)-(1→4)-O-(2,3,6-tri-O-benzyl-β-D-galactopyranosyl)-(1→4)-(3,6-di-**O-benzyl-2-O-pivaloyl-β-D-glucopyranoside** (6a). To a cooled (-10 °C) suspension of 9e (100 mg, 0.11 mmol) and 10 (160 mg, 0.17 mmol) in dry Et₂O (0.2 mL) was added TMSOTf (0.01 M in Et2O, 11 μ L) under an argon atmosphere. After stirring for 1 h, NaHCO₃ (s, 10 mg) and Et₂O were added, and the mixture was washed with saturated NaHCO₃ solution (5 mL) and H₂O (5 mL). The organic layer was dried (MgSO4), filtered, concentrated in vacuo, and the residue was purified by flash chromatography (9:1 toluene-EA) to afford 6a (119 mg, 62%): mp 87 °C; $[\alpha]^{rt}_{D}$ +36.7° (c 1, CHCl₃); TLC (7:1 toluene-acetone) $R_f 0.36$; ¹H-NMR δ 8.45 (s, 1H), 7.6–7.1 (m, 45H), 5.40 (s, 1H), 5.39 (s, 1H), 5.17 (d, J = 2.9 Hz, 1H), 5.06 (dd, J = 7.9, 9.5 Hz, 1H), 4.90–3.20 (m, 40H), 1.11 (s, 9H). Anal. Calcd for $C_{94}H_{99}N_4O_{21}Cl_3$: C, 65.37; H, 5.78; N, 3.24. Found: C, 65.29; H, 5.82; N, 3.44.

3-O-Acetyl-2-azido-4,6-O-benzylidene-2-deoxy-α- and -β-D-galactopyranosyl Trichloroacetimidate (7c). To a solution of **7b**^{24a} (9.3 g, 20.7 mmol) and AcOH (1.31 mL, 22.8 mmol) in dry THF (120 mL) at -20 °C was added dropwise TBAF (20.7 mL of a 1.1 M solution in THF). The mixture was then warmed up to 0 °C, diluted with Et₂O (500 mL), washed with saturated NaHCO₃ solution (300 mL) and brine (2×200 mL), dried (MgSO₄), and concentrated in vacuo. To a solution of this residue in dry CH₂Cl₂ (200 mL) was added Cl₃CCN (10 mL) and DBU (few drops). After stirring for 1 h, the solvent was removed in vacuo and the residue was purified by flash chromatography (8:1 toluene-acetone + 1% Et₃N) to yield 7c (8.0 g, 81%) as a 1:3.5 α/β mixture: TLC (5:1 toluene–acetone) R_f , 0.60, ¹H NMR: δ 8.74 and 8.72 (2 s, 1H), 7.6–7.3 (m, 5H), 6.58 (d, J = 2.9 Hz, 0.22H), 5.67 (d, J = 8.2 Hz, 0.78H), 5.52 and 5.51 (2 s, 1H), 5.31 (dd, J = 3.5, 9.6 Hz, 0.22H), 4.81 (dd, J = 3.6, 9.6 Hz, 0.78H), 2.17 and 2.15 (2 s, 3H). Anal. Calcd for C17H17N4O6Cl3: C, 42.56; H, 3.57; N, 11.68. Found: C, 42.30; H, 3.41; N, 11.76.

Allyl 3-*O*-(3-*O*-Acetyl-2-azido-4,6-*O*-benzylidene-2-deoxy- β -D-galactopyranosyl)-4,6-*O*-benzylidene- β -D-galactopyranoside (9f). To a suspension of 7c (7.50 g, 15.6 mmol) and 8a (5.78 g, 18.8 mmol) in dry CH₃CN (mL) was added TMSOTf (7.8 mL of a 0.1M solution in dry CH₃CN) at -20 °C under an argon atmosphere. After stirring for 2 h, the resulting clear solution was neutralized with NaHCO₃ (s, 1 g), filtered, and concentrated *in vacuo*. The residue was purified by flash chromatography (5:1 toluene-acetone) to give **9f** (8.22 g, 84%): [α]^{rt}_D +28.0° (*c* 1, CHCl₃); TLC (3:1 toluene-acetone) *R*_f 0.45, ¹H NMR: δ 7.6–7.2 (m, 10H), 5.94 (m, 1H), 5.57 (s, 1H), 5.48 (s, 1H), 5.35–5.15 (m, 2H), 4.96 (d, J = 8.0 Hz, 1H), 4.71 (dd, J = 3.5, 8.9 Hz, 1H), 4.5–3.99 (m, 11H), 3.85 (dd, J = 3.5, 9.9 Hz, 1H), 3.46 (br s, 1H), 3.41 (br s, 1H), 2.12 (s, 3H). Anal. Calcd for $C_{31}H_{35}N_{3}O_{11}$ ·¹/₂H₂O: C, 58.67; H, 5.71; N, 6.62. Found: C, 58.67; H, 5.68; N, 6.15.

Allyl 3-O-(3-O-Acetyl-2-azido-4,6-O-benzylidene-2-deoxy- β -D-galactopyranosyl)-2-O-benzyl-4,6-O-benzylidene- β -Dgalactopyranoside (9g). To a suspension of 9f (6.51 g, 10.4 mmol), freshly prepared Ag₂O (1.8 g), and 4 Å molecular sieves in dry DMF (50 mL) was added BnBr (1.8 mL, 15.6 mmol), and the mixture was stirred for 12 h at rt, filtered through Celite, and concentrated. The resulting residue was purified by column chromatography (7:1 toluene-acetone) to yield 9g (6.03 g, 81%): $[\alpha]^{rt}_{D} + 34.6^{\circ}$ (c 1, CHCl₃); TLC (4:1 toluene acetone) Rf 0.51, ¹H NMR: δ 7.6–7.2 (m, 15H), 5.96 (m, 1H), 5.59 (s, 1H), 5.47 (s, 1H), 5.38–5.15 (m, 2H), 4.99 (d, J = 9.8Hz, 1H), 4.93 (d, J = 8.0 Hz, 1H), 4.74 (d, J = 9.8 Hz, 2H), 4.63 (dd, J = 3.5, 10.9 Hz, 1H), 4.5–3.9 (m, 11H), 3.38 (br s, 1H), 3.32 (br s, 1H), 2.15 (s, 3H). Anal. Calcd for C₃₈H₄₁N₃-O₁₁: C, 63.77; H, 5.77; N, 5.87. Found: C, 63.78; H, 5.84; N, 5.87.

3-O-(3-O-Acetyl-2-azido-4,6-O-benzylidene-2-deoxy-β-Dgalactopyranosyl)-2-O-benzyl-4,6-O-benzylidene- α , β -Dgalactopyranose (9h). A mixture of compound 9g (5.00 g, 6.98 mmol), NaOAc (7.5 g, 83 mmol), and PdCl₂ (2.48 g, 14.0 mmol) in 9:1 AcOH-H₂O (100 mL) was stirred for 12h at 4 °C and then filtered through Celite. The filtrate was poured on ice-water (500 mL), extracted with CH_2Cl_2 (3 \times 200 mL), washed with saturated NaHCO₃ solution (2 \times 200 mL), and dried (MgSO₄). After removal of the solvent *in vacuo*, the resulting residue was purified by flash chromatography (7:1 toluene–acetone) to give **9h** (4.08 g, 86%): $[\alpha]^{rt}_{D} + 63.2^{\circ}$ (c 1, CHCl₃); TLC (4:1 toluene-acetone) *R*_f 0.27 and 0.18, ¹H NMR: δ 7.6–7.2 (m, 15H), 5.58 and 5.49 (2 s, 1H), 5.60 and 5.48 (2 s, 1H), 5.35 (d, J = 3.3 Hz, 1H), 5.0-3.9 (m, 16H), 3.43 (br s, 1H), 2.15 (s, 3H). Anal. Calcd for C₃₅H₃₇N₃O₁₁·1/2H₂O: C, 61.39; H, 5.59; N, 6.14. Found: C, 61.57; H 5.69; N, 6.11.

3-*O*-(**3**-*O*-Acetyl-2-azido-4,6-*O*-benzylidene-2-deoxy-β-Dgalactopyranosyl)-2-*O*-benzyl-4,6-*O*-benzylidene-α- and β-D-galactopyranosyl Trichloroacetimidate (9iα and 9iβ). To a solution of **9h** (2.60 g, 3.85 mmol) in dry CH₂Cl₂ (50 mL) was added Cl₃CCN (2 mL) and DBU (1 drop). After stirring for 1 h, the solvent was removed *in vacuo* and the residue was purified by flash chromatography (5:2 PE-EA + 1% Et3N). **9i**α eluted first (2.71 g, 86%): $[\alpha]^{rt}_D$ +92.4° (*c* 1, CHCl₃); TLC (5:2 PE-EA) *R*_f0.42; ¹H NMR: δ 8.58 (s, 1H), 7.6-7.2 (m, 15H), 6.64 (br s, 1H), 5.61 (s, 1H), 5.49 (s, 1H), 4.91 (d, *J* = 8.0 Hz, 1H), 4.72 (d, *J* = 10.9 Hz, 1H), 4.67 (d, *J* = 10.9 Hz, 1H), 4.62 (dd, *J* = 3.5, 10.8 Hz, 1H), 4.52 (br s, 1H), 4.4-3.9 (m, 9H), 3.39 (br s, 1H), 2.13 (s, 3H). Anal. Calcd for C₃₇H₃₇N₄O₁₁Cl₃: C, 54.19; H, 4.55; N, 6.83. Found: C, 54.02; H, 4.41; N, 6.87. **9i**β eluted second (0.22 g, 7%): $[\alpha]^{rt}_D$ +40.8° (*c* 1, CHCl₃);

9i β eluted second (0.22 g, 7%): $[\alpha]^{rt}_{D}$ +40.8° (*c* 1, CHCl₃); TLC R_f (5:2 PE-EA) R_f 0.30, ¹H NMR: δ 8.67 (s, 1H), 7.6-7.2 (m, 15H), 5.81 (d, J = 8.1 Hz, 1H), 5.61 (s, 1H), 5.49 (s, 1H), 4.99 (d, J = 9.9 Hz, 1H), 4.91 (d, J = 8.0 Hz, 1H), 4.79 (d, J = 9.9 Hz, 1H), 4.63 (dd, J = 3.5, 10.9 Hz, 1H), 4.4-3.2 (m, 40H), 2.15 (s, 3H). Anal. Calcd for C₃₇H₃₇N₄O₁₁Cl₃: C, 54.19; H, 4.55; N, 6.83. Found: C, 54.23; H, 4.42; N, 6.92.

Benzyl O-(3-O-Acetyl-2-azido-4,6-O-benzylidene-2-deoxy-β-D-galactopyranosyl)-(1→3)-O-(2-O-benzyl-4,6-O-benzylidene- α - and - β -D-galactopyranosyl)-(1 \rightarrow 4)-O-(2,3,6-tri- \hat{O} -benzyl- β -D-galactopyranosyl)-(1 \rightarrow 4)-2,6-di-O-benzyl- β -**D-glucopyranoside (6ca and 6c\beta).** To a solution of **10** (2 g, 2.1 mmol) in dry Et₂O (5 mL) was added TMSOTf (0.5 mL of a 0.1 M solution in dry Et₂O) under an argon atmosphere. A solution of **9i** α/β (2.5 g of a 12:1 α/β mixture, 3.1 mmol) in dry Et₂O (10 mL) was then added dropwise while the reaction stirred. After 30 min the mixture was neutralized with NaHCO₃ (s, 1 g), filtered, and concentrated in vacuo. The residue was purified by flash chromatography (4:1 PE-EA). **6c**α eluted first (1.71 g, 51%): [α]^{rt}_D +31.4° (c 1, CHCl₃); TLC (3:1 PE-EA) $R_f 0.47$, ¹H NMR: δ 7.5-7.0 (m, 45H), 5.43 (s, 1H), 5.35 (s, 1H), 5.20 (d, J = 2.7 Hz, 1H), 5.06 (dd, J = 8.0, 9.5 Hz, 1H), 4.9-3.2 (m, 40H), 2.11 (s, 3H), 1.11 (s, 9H). Anal. Calcd for C₉₄H₁₀₁N₃O₂₂: C, 69.48; H, 6.27; N, 2.59. Found: C, 69.77; H, 6.27; N, 2.84.

6cβ eluted second (0.40 g, 12%): $[\alpha]^{rt}_D$ +15.3° (*c* 1, CHCl₃); TLC (4:1 toluene–acetone) R_f 0.52, ¹H NMR: δ 7.7–6.7 (m, 45H), 5.48 (s, 1H), 5.16 (s, 1H), 5.13 (d, J = 10.2Hz, 1H), 5.06 (dd, J = 8.1, 9.5 Hz, 1H), 4.95–3.35 (m, 40H), 2.13 (s, 3H), 1.13 (s, 9H). Anal. Calcd for C₉₄H₁₀₁N₃O₂₂: C, 69.48; H, 6.27; N, 2.59. Found: C, 69.36; H, 6.35; N, 2.44.

Benzyl O-(2-Azido-4,6-O-benzylidene-2-deoxy-β-D-galactopyranosyl)-(1→3)-O-(2-O-benzyl-4,6-O-benzylideneα-D-galactopyranosyl)-(1→4)-O-(2,3,6-tri-O-benzyl-β-D-galactopyranosyl)-(1→4)-3,6-di-O-benzyl-2-O-pivaloyl-β-Dglucopyranoside (6b). (a) To a solution of 6a (100 mg, 57.9 µmol) in MeOH (5 mL) was added 1 N HCl (1 mL), and the mixture was stirred for 24 h at rt. After neutralization (saturated NaHCO₃ solution), the solvent was removed in vacuo and the residue extracted with acetone and purified by flash chromatography (3:1 PE-EA) to give **6b** (47 mg, 61%) as an amorphous powder: $[\alpha]^{rt}_{D}$ +21.7° (*c* 1, CHCl₃); TLC (2:1 PE-EA) $R_f 0.35$; ¹H-NMR δ 7.0–7.5 (m, 45H), 5.44 (s, 1H), 5.39 (s, 1H), 5.2 (d, J = 2.5 Hz, 1H), 5.08 (dd, J = 7.9, 9.6 Hz, 1H), 4.90-3.00 (m, 40H), 2.42 (d, J = 9.7 Hz, 1H), 1.12 (s, 9H). Anal. Calcd for $C_{92}H_{99}N_3O_{21}$: C, 69.81; H, 6.30; N, 2.65. Found: C, 69.83; H, 6.20; N, 2.87.

(b) To a solution of $6c\alpha$ (1.65 g, 1.02 mmol) in dry MeOH (5 mL) was added saturated methanolic ammonia solution (10 mL), and the mixture was stirred for 6 h at rt. After removal of the solvent, the residue was purified through a short silica gel column (2:1 PE-EA) to yield **6b** (1.61 g, quant), identical with the product described above.

Benzyl O-(2,3-Di-O-Acetyl-4,6-O-benzylidene-β-D-galactopyranosyl)-(1→3)-O-(2-azido-4,6-O-benzylidene-2-deoxy- β -D-galactopyranosyl)-(1 \rightarrow 3)-O-(2-O-benzyl-4,6-O-benzylidene-α-D-galactopyranosyl)-(1→4)-O-(2,3,6-tri-O-benzyl- β -D-galactopyranosyl)-(1 \rightarrow 4)-3,6-di-O-benzyl-2-O-pivaloyl- β -**D**-glucopyranoside (3a). To a stirred suspension of **6b** (1.5 g, 0.95 mmol) and 5 (0.71 g, 1.42 mmol) in dry Et₂O (5 mL) was added TMSOTf (0.47 mL of a 0.1 M solution in dry Et₂O) under an argon atmosphere. The mixture was stirred for 30 min at rt, $NaHCO_3$ (s, 1 g) was added, and the mixture was filtered and concentrated *in vacuo*. The residue was purified by flash chromatography (2:1 PE-EA) to yield 3a (1.82 g, 94%): $[\alpha]^{rt}_{D}$ +44.8° (*c* 1, CHCl₃); TLC (7:4 PE-EA) R_f 0.22, ¹H NMR: δ 7.6–7.0 (m, 50H), 5.53 (s, 1H), 5.44 (s, 1H), 5.41 (s, 1H), 5.39 (dd, J = 8.0, 10.4 Hz, 1H), 5.18 (d, J = 2.9 Hz, 1H), 5.09 (dd, J = 8.0, 9.5 Hz, 1H), 4.99 (dd, J = 3.6, 10.4 Hz, 1H), 4.9-3.15 (m, 45H), 2.08 (s, 3H), 2.03 (s, 3H), 1.13 (s, 9H). Anal. Calcd for C109H117N3O28: C, 68.29; H, 6.15; N, 2.19. Found: C, 67.88; H, 6.27; N, 2.37.

Benzyl *O*-(4,6-*O*-Benzylidene-β-D-galactopyranosyl)-(1→3)-O-(2-azido-4,6-O-benzylidene-2-deoxy-β-D-galactopyranosyl)-(1→3)-O-(2-O-benzyl-4,6-O-benzylidene-α-Dgalactopyranosyl)-(1→4)-O-(2,3,6-tri-O-benzyl-β-D-galactopyranosyl)-(1→4)-3,6-di-O-benzyl-2-O-pivaloyl-β-Dglucopyranoside (3b). To a solution of 3a (533 mg, 0.28 mmol) in dry MeOH (3 mL) was added saturated methanolic ammonia solution (5 mL) and the mixture was stirred for 4 h at rt. After removal of the solvent, the residue was purified through a short silica gel column (4:1 toluene-acetone) to yield **3b** (510 mg, quant): $[\alpha]^{rt}_{D}$ + 48.0° (c 1, CHCl₃); TLC (4:1 toluene-acetone) R_f 0.30, ¹H NMR: δ 7.6–7.0 (m, 50H), 5.58 (s, 1H), 5.46 (s, 1H), 5.45 (s, 1H), 5.20 (d, J = 3.0 Hz, 1H), 5.10 (dd, J = 8.0, 9.6 Hz, 1H), 4.9–3.1 (m, 47H), 2.83 (br s, 1H), 2.52 (br s, 1H), 1.13 (s, 9H). Anal. Calcd for C₁₀₅H₁₁₃N₃O₂₆: C, 68.80; H, 6.21; N, 2.29. Found: C, 68.56; H, 6.30; N, 2.40.

Benzyl O-(Methyl 5-Acetamido-4,7,8,9-tetra-O-acetyl-3,5-dideoxy-D-glycero- α - and β -D-galacto-2-nonulopyranosylonate)-(2 \rightarrow 3)-O-(4,6-O-benzylidene- β -D-galactopyranosyl)-(1 \rightarrow 3)-O-(2-azido-4,6-O-benzylidene-2-deoxy- β -Dgalactopyranosyl)-(1 \rightarrow 4)-O-(2,3,6-tri-O-benzylidene- α -D-galactopyranosyl)-(1 \rightarrow 4)-O-(2,3,6-tri-O-benzyl- β -Dgalactopyranosyl)-(1 \rightarrow 4)-3,6-di-O-benzyl-2-O-pivaloyl- β -D-glucopyranoside (11a and 11a β). To a solution of 3b (300 mg, 0.16 mmol of a 5:1 α/β mixture) and 2 (150 mg, 0.25 mmol) in dry CH₃CN (3 mL) was added TMSOTf (0.16 mL of a 0.1 M solution in dry CH₃CN) at -50 °C under an argon atmosphere. After stirring for 30 min, more 2 (50 mg, 0.08 mmol, solution in 0.5 mL of dry CH₃CN) was added and the mixture stirred for another 30 min. After addition of NaHCO₃ (s, 0.1 g), the reaction mixture was filtered and concentrated *in vacuo*, and the resulting residue was purified by flash chromatography (3:1 toluene–acetone). **11a** β eluted first (49 mg, 13%): [α]^{rt}_D +23.1° (*c* 1, CHCl₃); TLC (4:1 toluene–acetone) R_f 0.40, ¹H NMR: δ 7.0–7.7 (m, 35H), 5.72 (s, 1H), 5.44 (s, 1H), 5.42 (s, 1H), 5.41 (m, 1H), 5.35 (m, 1H), 5.30 (m, 1H), 5.26 (dd, J = 2.3, 12.2 Hz, 1H), 5.18 (d, J = 3.0 Hz, 1H), 5.09 (dd, J = 7.9, 9.4 Hz, 1H), 4.66 (d, J = 7.7 Hz, 1H), 4.58 (d, J = 8.6 Hz, 1H), 4.44 (d, J = 8.1 Hz, 1H), 3.97 (m, 1H), 3.78 (s, 3H), 2.74 (br s, 1H), 2.67 (dd, J = 4.4, 12.2 Hz, 1H), 2.14 (s, 3H), 2.08 (s, 3H), 2.03 (s, 3H), 2.00 (s, 3H), 1.90 (t, J = 13.1 Hz, 1H), 1.67 (s, 3H), 1.13 (s, 9H). Anal. Calcd for C₁₂₅H₁₄₀N₄O₃₈: C, 65.09; H, 6.12; N, 2.43. Found: C, 64.88; H 6.28; N, 2.64.

11a α eluted second (180 mg, 48%): $[\alpha]^{rt_D} + 24.6^{\circ}$ (*c* 1, CHCl₃), ¹H NMR: δ 7.5-7.0 (m, 50H), 5.45 (s, 2H), 5.40 (s, 1H), 5.43 (m, 1H), 5.30 (dd, J = 1.6, 9.2 Hz, 1H), 5.17 (d, J = 3.2 Hz, 1H), 5.16 (d, J = 8.7 Hz, 1H), 5.09 (dd, J = 8.0, 9.4 Hz, 1H), 4.88 (m, 1H), 4.66 (d, J = 7.7 Hz, 1H), 4.58 (d, J = 8.8 Hz, 1H), 4.48 (d, J = 8.6 Hz, 1H), 4.44 (d, J = 8.1 Hz, 1H), 4.35 (m, 1H), 4.02 (m, 1H), 3.50 (s, 3H), 2.74 (dd, J = 4.5, 12.7 Hz, 1H), 2.73 (br s, 1H), 2.21 (s, 3H), 2.16 (s, 3H), 2.01 (s, 3H), 1.99 (s, 3H), 2.03 (t, J = 12.8 Hz, 1H), 1.88 (s, 3H), 1.13 (s, 9H). Anal. Calcd for $C_{125}H_{140}N_4O_{38}$: C, 65.09; H, 6.12; N, 2.43. Found: C, 65.00; H 6.39; N, 3.05.

O-(Methyl 5-Acetamido-4,7,8,9-tetra-O-acetyl-3,5-dideoxy-D-glycero- α -D-galacto-2-nonulopyranosylonate)-(2 \rightarrow 3)-O-(2,4,6-tri-O-acetyl- β -D-galactopyranosyl)-(1 \rightarrow 3)-O-(2acetamido-4,6-di-O-acetyl-2-deoxy-β-D-galactopyranosyl)- $(1\rightarrow 3)$ -O-(2,4,6-tri-O-acetyl- α -D-galactopyranosyl)- $(1\rightarrow 4)$ -O-(2,3,6-tri-O-acetyl- β -D-galactopyranosyl)-(1 \rightarrow 4)-1,3,6tri-*O*-acetyl-2-*O*-pivaloyl- α/β -D-glucopyranoside (11b). A mixture of 11a (145 mg, 62.8 mmol) and freshly prepared³² Pd(OH)₂ (19% on carbon, 50 mg) in MeOH (50 mL) was stirred under H₂ (4 bar) at rt for 24 h. The mixture was filtered and washed with MeOH (50 mL). The filtrate was concentrated, and the resulting residue was coevaporated with toluene (2 \times 10 mL), dissolved in 1:1 Py-Ac₂O (3 mL), and kept at rt for 24 h. The mixture was then poured on ice-water (50 mL) and extracted with CH_2Cl_2 (3 \times 25 mL). The organic layer was washed with 1 M HCl (50 mL), saturated NaHCO₃ solution (50 mL), and brine (50 mL). After removal of the solvent in vacuo, the resulting residue was purified by flash chromatography (40:1 CHCl₃–MeOH) to yield **11b** (108 mg, 85%) as a 1:1 mixture of α and β anomers: TLC (30:1 CHCl₃–MeOH) $R_f 0.29$, ¹H NMR: δ 6.30 (d, J = 3.7 Hz), 4.05 (m, 1H), 3.85 (s, 3H), 2.58 (dd, J = 4.6, 12.7 Hz, 1H), 2.25-1.80 (m, 60H), 1.68 (t, J = 12.4 Hz, 1H), 1.13 and 1.14 (2 s, 9H); MS-FAB (thioglycerol matrix, NaI) m/z 2037 (MNa⁺). Anal. Calcd for C₈₅H₁₁₈N₂O₅₃: C, 50.64; H, 5.90; N, 1.39. Found: C, 50.70; H 6.07; N. 1.01

O-(Methyl 5-Acetamido-4,7,8,9-tetra-O-acetyl-3,5-dideoxy-D-glycero- α -D-galacto-2-nonulopyranosylonate)-(2 \rightarrow 3)-O-(2,4,6-tri-O-acetyl- β -D-galactopyranosyl)-(1 \rightarrow 3)-O-(2acetamido-4,6-di-O-acetyl-2-deoxy-β-D-galactopyranosyl)-(1→3)-O-(2,4,6-tri-O-acetyl-α-D-galactopyranosyl)-(1→4)-O-(2,3,6-tri-O-acetyl-β-D-galactopyranosyl)-(1→4)-3,6-di-**O-acetyl-2-O-pivaloyl-** α , β -D-glucopyranose (11c). A solution of 11b (100 mg, 49.7 μ mol) in dry DMF (3 mL) was stirred with NH₂NH₂·AcOH (8 mg) at 40 °C for 1 h. The mixture was then diluted with AcOEt (30 mL) and washed with brine (20 mL). The aqueous layer was washed (AcOEt, 2×15 mL) and the combined organic layer was concentrated in vacuo and purified by column chromatography (40:1 CH₂Cl₂-MeOH) to yield **11c** (92 mg, 94%): $[\alpha]^{rt}_D$ +55.6° (*c* 1, CHCl₃); TLC (20:1 CHCl₃-MeOH) R_f 0.44, ¹H NMR: 6.18 (d, J = 8.0 Hz, 1H), 3.81 (s, 3H), 2.54 (dd, J = 4.1, 12.2 Hz, 1H), 2.25–1.82 (m, 57H), 1.67 (t, J = 12.3 Hz, 1H), 1.15 (s, 9H); MS-FAB (thioglycerol matrix, NaI) m/z 1995 (MNa⁺). Anal. Calcd for $C_{83}H_{116}N_2O_{52}$: C, 50.50; H, 5.92; N, 1.42. Found: C, 50.22; H 6.11; N, 1.25

O-(Methyl 5-Acetamido-4,7,8,9-tetra-O-acetyl-3,5-dideoxy-D-glycero-α-D-galacto-2-nonulopyranosylonate)- $(2\rightarrow 3)$ -O-(2,4,6-tri-O-acetyl-β-D-galactopyranosyl)- $(1\rightarrow 3)$ -O-(2acetamido-4,6-di-O-acetyl-2-deoxy-β-D-galactopyranosyl)- (1→3)-*O*-(2,4,6-tri-*O*-acetyl-α-D-galactopyranosyl)-(1→4)-*O*-(2,3,6-tri-*O*-acetyl-β-D-galactopyranosyl)-(1→4)-3,6-di-*O*-acetyl-2-*O*-pivaloyl-α-D-glucopyranosyl Trichloroacetimidate (11d). To a solution of 11c (80 mg, 40 µmol) in dry CH₂Cl₂ (5 mL) was added Cl₃CCN (40 µL) and DBU (1 drop). After stirring for 2 h, the solvent was removed *in vacuo* and the residue was purified through a short silica gel column (50:1 CH₂Cl₂-MeOH) to afford 11d (80 mg, 93%): [α]⁺_D+72.8° (*c* 1, CHCl₃); TLC (40:1 CH₂Cl₂-MeOH) *R*_ℓ 0.28, ⁺H NMR: δ 8.66 (s, 1H), 6.51 (d, *J* = 3.6 Hz, 1H), 3.84 (s, 3H), 2.58 (dd, *J* = 4.6, 12.9 Hz, 1H), 2.25-1.94 (m, 54H), 1.85 (s, 3H), 1.73 (s, 3H), 1.68 (t, *J* = 12.3 Hz, 1H), 1.14 (s, 9H). Anal. Calcd for C₈₅H₁₁₆N₄O₅₂Cl₃: C, 48.20; H, 5.52; N, 2.65. Found: C, 48.13; H, 5.61; N, 2.43.

O-(Methyl 5-Acetamido-4,7,8,9-tetra-O-acetyl-3,5-dideoxy-D-glycero-α-D-galacto-2-nonulopyranosylonate)-(2→3)-O-(2,4,6-tri-O-acetyl- β -D-galactopyranosyl)-(1 \rightarrow 3)-O-(2acetamido-4,6-di-O-acetyl-2-deoxy-β-D-galactopyranosyl)- $(1\rightarrow 3)$ -O-(2,4,6-tri-O-acetyl- α -D-galactopyranosyl)- $(1\rightarrow 4)$ -O-(2,3,6-tri-O-acetyl-β-D-galactopyranosyl)-(1→4)-3,6-di-*O*-acetyl-2-*O*-pivaloyl- β -D-glucopyranosyl-(1 \rightarrow 1)-(2*S*,-3R,4E)-2-azido-3-O-benzoyl-4-octadecene-1,3-diol (12a). To a solution of **11d** (70 mg, 30 µmol) and (2*S*,3*R*,4*E*)-2-azido-3-(benzoyloxy)-4-octadecen-1-ol (4) (25 mg, 60 µmol) in dry CH2- Cl_2 (200 µL) was added TMSOTf (17 µL of a 0.01 M solution in dry CH₂Cl₂) under an argon atmosphere and the mixture was stirred at rt for 1 h. NaHCO₃ (s, 10 mg) was then added and the mixture was filtered, concentrated, and purified by flash chromatography (40:1 CH₂Cl₂-MeOH) to yield 23 (57 mg, 72%): $[\alpha]^{rt}_{D} + 29.7^{\circ}$ (c 1, \tilde{CHCl}_{3}); TLC (25:1 CH₂Cl₂-MeOH) $R_f 0.45$, ¹H NMR: δ 8.1–7.4 (m, 5H), 5.91 (dt, J = 6.5, 14.5, 1H), 5.75 (d, J = 7.4 Hz, 1H), 4.70 (d, J = 8.0 Hz, 1H), 3.85 (s, 3H), 3.34 (m, 1H), 2.59 (dd, J = 4.3, 12.4 Hz, 1H), 2.25 -1.94 (m, 53H), 1.85 (s, 3H), 1.80 (s, 3H), 1.68 (t, J = 12.5 Hz, 1H), 1.4–1.2 (m, 22H), 1.17 (s, 9H), 0.88 (t, J = 6.6 Hz, 3H); MS-FAB (thioglycerol matrix + NaI) m/z 2406 (MNa⁺). Anal. Calcd for $C_{108}H_{153}N_5O_{54}$: C, 54.38; H, 6.46; N, 2.94. Found: C, 54.66; H 6.47; N, 2.77.

O-(Methyl 5-Acetamido-4,7,8,9-tetra-O-acetyl-3,5-dideoxy-D-glycero- α -D-galacto-2-nonulopyranosylonate)-(2 \rightarrow 3)-O-(2,4,6-tri-O-acetyl- β -D-galactopyranosyl)-(1 \rightarrow 3)-O-(2acetamido-4,6-di-O-acetyl-2-deoxy- β -D-galactopyranosyl)- $(1\rightarrow 3)$ -O-(2,4,6-tri-O-acetyl- α -D-galactopyranosyl)- $(1\rightarrow 4)$ -O-(2,3,6-tri-O-acetyl- β -D-galactopyranosyl)-(1 \rightarrow 4)-3,6-di-*O*-acetyl-2-*O*-pivaloyl- α , β -D-glucopyranosyl-(1 \rightarrow 1)-(2*S*,-3R,4E)-2-hexadecanamido-3-O-benzoyl-4-octadecene-1,3diol (12b). H_2S was slowly bubbled through a solution of 23 (35 mg, 14.7 μ mol) in 8:2 Py-H₂O (4 mL) for 2 h at rt. After stirring for 48 h, the solvent was concentrated in vacuo, the bath temperature being kept under 30 °C. To a solution of the residue in dry CH_2Cl_2 (3 mL) were added palmitic acid (9 mg, 34 μ mol) and N-(3-(N,N-dimethylamino)propyl)-N-ethylcarbodiimide hydrochloride (WSC) (9 mg, 46 μ mol), and the mixture was stirred for 18 h at rt. The reaction mixture was then diluted with more CH₂Cl₂ (15 mL), washed with H₂O (2 imes 5 mL), and concentrated *in vacuo*. The residue was purified by flash chromatography (40:1 CH₂Cl₂-MeOH) to yield 12b (33 mg, 87%): $[\alpha]^{rt}_{D} + 32.9^{\circ}$ (*c* 1, CHCl₃); TLC (25:1 CH₂Cl₂– MeOH) R_{ℓ} 0.50, ¹H NMR: δ 8.1–7.4 (m, 5H, Ph), 5.88 (dt, J =6.6, 14.7, 1H, H-5cer), 5.75 (d, J = 8.2 Hz, 1H, NH-2d), 5.73 (d, J = 9.8 Hz, 1H, NH-2cer), 5.6–5.4 (m, 5H, H-8f, H-3cer, H-1c, H-4d, H-4cer), 5.37 (dd, J = 2.5, 8.8 Hz, 1H, H-7f), 5.18 (d, J = 8.7 Hz, 1H, H-1d), 4.69 (d, J = 7.8 Hz, 1H, H-1e), 4.48 (m, H-2cer), 4.47 (d, J = 8.3 Hz, 1H, H-1b), 4.44 (d, J = 8.0 Hz, 1H, H-1a), 3.85 (s, 3H, CO₂Me), 2.59 (dd, J = 4.5, 12.5 Hz, 1H, H-3feq), 2.25–1.90 (m, 53H, 17 OAc, H-6cer, H-6'cer), 3.33 (m, 1H, H-2d), 1.85 (s, 3H, NAc), 1.73 (s, 3H, NAc), 1.15 (s, 9H, OPiv), 0.88 (t, J = 6.5 Hz, 6H, 2 CH₃); MS-FAB (thioglycerol matrix, NaI) m/z 2620 (MNa⁺). Anal. Calcd for C₁₂₄H₁₈₆N₃O₅₅: C, 57.33; H, 7.18; N, 1.62. Found: C, 56.98; H 7.29; N, 1.45.

O-(5-Acetamido-3,5-dideoxy-D-glycero-α-D-galacto-2nonulopyranosylonic Acid)- $(2 \rightarrow 3) - O - (\beta - D - galactopyrano$ syl)-(1→3)-O-(2-acetamido-2-deoxy-β-D-galactopyranosyl)- $(1\rightarrow 3)$ -O- $(\alpha$ -D-galactopyranosyl)- $(1\rightarrow 4)$ -O- $(\beta$ -D-galactopyranosyl)- $(1\rightarrow 4)$ - β -D-glucopyranosyl- $(1\rightarrow 1)$ -(2S, 3R, 4E)-2hexadecanamido-4-octadecene-1,3-diol (1). To a solution of 12b (22 mg, 8.5 μ mol) in dry MeOH (1.5 mL) was added NaOMe (0.2 M in MeOH, 0.35 mL) and the mixture was stirred for 24 h at 40 °C. KOH (0.2 M in water, 0.4 mL) was added and the mixture stirred at rt for another 24 h. The solution was then neutralized with ion-exchange resin Amberlite IR 120 (H⁺), filtered, and concentrated *in vacuo*. The residue was purified by column chromatography (1:1 CH₂Cl₂-EtOH) on Sephadex LH-20 (15 mL) to yield 1 (14.2 mg, quantitative) as a white powder: $[\alpha]^{rt}_{D} + 8.5^{\circ}$ (*c* 0.4, Py); TLC (8:4:3 *n*-BuOH-AcOH $-\hat{H}_2$ O) R_f 0.37, ¹H NMR (500 MHz, 95:5 DMSO- d_6 /D₂O, 40 °C): δ 5.51 (dt, J = 6.8, 15.4, 1H, H-5cer), 5.33 (dd J = 7.1, 15.4 Hz, 1H, H-4cer), 4.79 (d, J = 4.0 Hz, 1H, H-1c), 4.56 (d, J = 7.3 Hz, 1H, H-1d), 4.24 (d, J = 7.7 Hz, 1H, H-1b or e), 4.22 (d, J = 7.9 Hz, 1H, H-1b or e), 4.16 (d, J = 7.8 Hz, 1H, H-1a), 3.86 (m, 1H, H-3cer), 3.53 (m, 1H, H-4f), 3.02 (dd J= 7.2, 8.2, 1H, H-2a), 2.74 (dd J = 4.6, 11.8, 1 H, H-3feq), 2.01 (t, J = 7.1 Hz, 2H, COCH₂), 1.91 (m, 2H, H-6cer, H-6'cer), 1.86 (s, 3H, NAc), 1.78 (s, 3H, NAc), 1.34 (t, J = 11.6 Hz, 1H, H-3fax), 0.83 (t, J = 6.9 Hz, 6H, 2 CH₃). MS-FAB (thioglycerol/ PNB matrix, NaI) m/z 1725 (M + 2Na⁺); (thioglycerol/PNB matrix, CsI) m/z 1945 (M + 2Cs⁺).

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Supporting Information Available: ¹H NMR spectrum for compound **1** (2 pages). This material is contained in libraries on microfiche, immediately follows this article in the microfilm version of the journal, and can be ordered from the ACS; see any current masthead page for ordering information.

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